This article was downloaded by: [Univ Politec Cat] On: 31 December 2011, At: 07:05 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Chemistry and Ecology

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/gche20

Speciation of heavy metals in sediments from the Scheldt estuary, Belgium

D. Shilla ^a & J. Dativa ^b

^a Department of Biology, University of Antwerp, Belgium

^b Department of Chemistry, Dar es Salaam University, Dar es Salaam, Tanzania

Available online: 16 Feb 2011

To cite this article: D. Shilla & J. Dativa (2011): Speciation of heavy metals in sediments from the Scheldt estuary, Belgium, Chemistry and Ecology, 27:1, 69-79

To link to this article: <u>http://dx.doi.org/10.1080/02757540.2010.529075</u>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <u>http://www.tandfonline.com/page/terms-and-conditions</u>

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



Taylor & Francis Taylor & Francis Group

Speciation of heavy metals in sediments from the Scheldt estuary, Belgium

D. Shilla^a* and J. Dativa^b

^aDepartment of Biology, University of Antwerp, Belgium; ^bDepartment of Chemistry, Dar es Salaam University, Dar es Salaam, Tanzania

(Received 4 June 2010; final version received 8 September 2010)

A simple three-step sequential extraction procedure was applied to study the speciation of heavy metals in sediment from Scheldt estuary, and their relationship to sediment grain size and organic matter content. The sedimentary metal content was fractionated into carbonate and exchangeable, metals bound to organic matter and residual fractions. Sedimentary total metal content was also determined using an industrial microwave (ETHOS 900) HF/HNO₃ extraction method. The extracts were analysed for metals using inductively coupled plasma atomic emission spectrometry. The bioavailable fraction (exchangeable and metals bound to organic matter) comprised less than the other forms. Residue metals were the dominant form of metals in almost all studied sites. The average total metal content for the studied sites, decreased in the order Fe > Cr > Cu > Co > Zn > Pb > Cd. Based on average values for the studied sites, the highest bioavailable metals in sediments were Cd (38%) from Westkapelle, Zn (17%) from Yerseke, Co (12%) from Domburg, Cr (9%) from Vlissingen, Fe and Pb each (2%) from Yerseke, and Cu (1%) from Domburg. Metal recovery was good, with <10% difference between the total metal recovered through the extractant steps and the total metal determined using HF/HNO₃ extract.

Keywords: heavy metals; Scheldt estuary; sediment; sequential extraction; speciation; bioavailable

1. Introduction

Total metal concentration has been exhausted in most studies to determine trace metals in sediment [1]. It is a good tool that gives some indication of the level of contamination in sediments, but consequently, the total metal concentration in sediments is not a reliable indicator of bioavailable metal, i.e. it does not give sufficient information about the mobility and bioavailability of metals [2–4]. Metals in sediments are associated with a number of physico-chemical forms, which in turn affect their bioavailability to organisms. Ray et al. [5] recommended that metals associated with an EDTA extract are a good measure of available metal. Alternatively, Rule and Alden [6] suggested that the sum of amounts of metals found in exchangeable and easily reducible fractions can be used to indicate Cd bioavailability to blue mussel *Mytilus edulis*. Fractionation of metals from sediments can therefore differentiate metal species and define fractions which are related to their specific nature [7].

ISSN 0275-7540 print/ISSN 1029-0370 online © 2011 Taylor & Francis DOI: 10.1080/02757540.2010.529075 http://www.informaworld.com

^{*}Corresponding author. Email: dshilla@yahoo.com

Depending on the toxic metals, there are quite large differences between their total and ecologically effective amounts. Because determination of the total metal gives little information about speciation, and hence the bioavailable fraction of metals to organisms, methods to determine different forms of metals in sediments have been proposed. In particular, a sequential extraction procedure has been widely used to achieve this goal (i.e. to speciate potentially toxic metals in a variety of samples) [8–10]. In a sequential extraction procedure, a series of reagents is used to extract operationally defined discrete phases from sediment in an outlined sequence [11]. The phases are usually adsorptive or exchangeable, carbonate, reducible (Fe-Mn oxide), organic and residue [12]. Many investigators have performed studies on the quantitative distribution of metals among various chemical phases of sediment with the aim of assessing the toxic potential of pollutants in the environment [13–16]. Fractionation is performed by a sequence of selective chemical leaching techniques that include the successive removal of these phases and their associated metals [12]. Tessier and others developed the first sequential extraction procedure in 1979 [17]. Since then, several attempts to modify the sequential extraction procedure for the assessment of metal speciation among different phases have been made, with different degrees of success, aimed at improving the selectivity of the extracting agent towards a particular phase and reducing the operation time to facilitate its routine application [18,19].

In our study, a simple sequential extraction procedure with few steps, but still relevant for metal bioavailability and accumulation, was used with the aim of studying the speciation of heavy metals in the sedimentary phase, and their relationship to the organic matter content and sediment grain size.

2. Materials and methods

2.1. The Scheldt estuary

Baeyens et al. [20] extensively described the Scheldt estuary. The River Scheldt (Figure 1) starts in the northern part of France (St Quentin) and empties into the North Sea near Vlissingen (The



Figure 1. Map showing location of the sampling sites. Stations with code numbers 1, 2, 3, 4, 5, 6 and 7 are Domburg, Westkapelle, Vlissingen, Borsele, Hansweert, Wemeldinge and Yerseke, respectively.

2.2. Sampling campaign

The locations of the sampling sites are presented in Figure 1. In total, seven sampling sites in the Scheldt estuary were selected and sampled along the salinity gradient. All sediment samples were collected during the winter season (30 January 2002 to 13 February 2002) at low tide. Surface (0-5 cm) sediment samples were collected at the edge of the water using a hand corer. Samples were put into clean 20 cm³ polyethylene bottles and tightly closed. Five replicates of the sediment samples were collected per site to study metal speciation on different sedimentary phases and for total metal analysis. The samples were put immediately into plastic bags, stored in an icebox, transferred to the laboratory and then stored in the freezer at -20 °C until all the moisture content had turned to ice. Samples were then taken into the freeze drier for lyophilisation. Thereafter, samples were analysed for heavy metals.

2.3. Sequential extraction procedure

The sequential extraction method was carried out in three steps, at each of which a specific extraction solution was added. All water used was of Milli-Q (Millipore; resistively $18 \text{ M} \cdot \text{cm}^{-1}$) quality. We applied a combined simple three-step sequential extraction procedure, partly adopted from Community Bureau of Reference (BCR) or Standards, Measurement and Testing (SMT) for fractions 1 and 3 (F₁ and F₃), and partly from Whitworth et al. [21] for fraction 2 (F₂) (Figure 2). The two fractions (F₁, carbonate and exchangeable metals and F₂, metals bound to organic matter) are considered to be the most labile or leachable and hence relevant for bioavailability and accumulation by aquatic organisms [4].



Figure 2. Schematic presentation of the sequential extraction protocol followed in this study.

In the first step, ~ 2 g of dry homogenised sediment sample was transferred into a pre-cleaned polycarbonate tube and 50 mL of 0.1 M acetic acid was added. The mixture was shaken for 16 h (overnight) in a horizontal end-over-end shaker at 140–150 rpm at room temperature. Care was taken to avoid any delay between the addition of the extracting solution and the starting of shaking. The solution was then centrifuged at 5000 rpm for 25 min and the supernatant fluid decanted into a pre-cleaned 60 mL HDPE bottle and stored at 4 °C until measurement with inductively coupled plasma mass spectrometry (ICP-MS) or inductively coupled plasma atomic emission spectrometry (ICP-AES).

Fifty millilitres of 0.05 M EDTA solution was added to the washed residue of step 1; the mixture was then shaken in an end-over-end shaker at 140–150 rpm for 72 h at room temperature. The solution was then centrifuged at 5000 rpm for 10 min and the supernatant fluid decanted into a pre-cleaned 60 mL HDPE bottle and stored at 4 °C until measurement with ICP-MS. The residue sediment from this extraction step was washed with 20 mL of milli-Q water, frozen, lyophilised and used for the third step, which is residue total metal extraction.

Total metal extraction from the sediment samples was performed using an industrial microwave (ETHOS 900) HF/HNO₃ extraction and the HF acid was neutralised by H₃BO₃ according to Van Ryssen et al.'s [22] total digestion protocol for soil or sediment samples. Approximately 0.2 g of dried and homogenised sediment samples were weighed in the digestion vessel. Four millilitres of sub-boiled (purified) HNO₃ and 4 mL of sub-boiled (purified) HF was added. The digestion pumps were closed tightly according to the instruction manual, and placed in the rotor inside the industrial microwave (ETHOS 900) and the digestion program was started as described by Van Ryssen et al. [22]. Blanks were analysed alongside samples and received the same treatment. The method was checked by analysing reference sediment samples (CRM 320). The values from the experiment and their standard deviations were in good agreement with values for the certified material (see Table 3).

Carbon content in sediment samples was analysed using a C:N analyser (Carlo Erba Strumentazione NA 1500 nitrogen analyser). Both total and organic carbon were analysed. The difference between total carbon and total organic carbon was recorded to represent inorganic carbon. To measure the organic carbon, sediment samples ($\sim 10 \text{ mg}$) were spiked with a few drops of 5 M HCl and placed in an oven at 50 °C for at least 1 h to dry. Decalcified samples were then analysed in the C:N analyser and the valued recorded. The quality of this analysis was controlled by analysing blanks (empty silver capsules) and a standard reference material (acetanilide). Sediment samples were analysed for grain sizes using a grain size analyser (Malvern Instruments Ltd, UK) and various size fractions were composed as recorded in Table 1.

All chemicals used were of analytical reagent grade from Merck Co. All materials in contact with the samples during sampling campaigns, all glass and plastic ware were subjected to decontamination processes by filling and storing them in 5–15% nitric acid (HNO₃, Pro analysis) at

Site code	Grain size fractions in (%)									
	$<2\mu m$	2–16 µm	16-63 µm	$63-125\mu m$	$125250\mu\text{m}$	$250500\mu\text{m}$	500–1000 μm			
1	0.32 (0.02)	0.05 (0.02)	0.03 (0.01)	0.07 (0.01)	7.53 (0.77)	73.93 (9.95)	18.08 (10.70)			
2	0.33 (0.22)	0.08 (0.15)	0.00 (0.00)	0.74 (0.14)	33.67 (3.19)	64.61 (2.61)	0.58 (0.41)			
3	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.30 (0.11)	18.79 (3.31)	72.69 (0.91)	8.22 (2.58)			
4	1.96 (1.51)	4.57 (4.43)	13.60 (2.69)	18.48 (6.29)	37.34 (2.37)	23.25 (13.00)	0.80 (0.82)			
5	6.57 (0.81)	20.02 (2.67)	40.18 (1.69)	20.55 (2.23)	10.80 (2.27)	1.90 (0.86)	0.00 (0.00)			
6	0.67 (0.06)	0.49 (0.13)	1.30 (0.50)	16.96 (5.70)	49.33 (7.51)	25.02 (9.57)	6.22 (3.60)			
7	1.10 (0.09)	1.07 (0.06)	2.57 (0.37)	21.14 (2.14)	60.71 (1.45)	13.41 (1.39)	0.00 (0.00)			

Table 1. Mean (n = 5) percentage volume of the sediment grain size fractions (μm) used in the study.

Note: Numbers in parentheses represent percentage standard deviations.

room temperature for at least 24 h prior to use, rinsing at least three times with Milli-Q-water (Millipore; resistively $18 \,\mathrm{M} \cdot \mathrm{cm}^{-1}$) quality.

2.4. Statistical analysis

Correlation analysis was made to test for a relationship between heavy metals and both carbon content and sediment grain size. The strength and significance of the parameters estimated was evaluated using correlation coefficients (r) and significance levels (p). All statistical analyses were performed using SPSS software version 11.

3. Results

3.1. Grain size distribution and the carbon content of the sediment

The percentage distribution of different size fractions in sediments from sites 1–5 (west Scheldt), 6 and 7 (eastern Scheldt) which were used in the speciation study are given in Table 1. The average content of the < 63 μ m fraction was 24.74% by volume with a minimum of 0.30% and a maximum of 87.32% by volume. In the < 63 μ m sediment fraction, the average proportion of clay-size sediment (< 4 μ m) ranged from 0.00 to 26.59% by volume. Sediments with a clay content > 5% were found mainly at site 5. The distribution of organic carbon (C_{org}) was similar to grain size distribution for all sites. The inorganic carbon content in the sediment was inverse to that of organic carbon. The carbon content distribution in the sediment is shown in Table 2.

Organic carbon in marine sediments may either be terrigenous or derived from biological productivity in the marine environment. Higher primary and secondary productivities in marine environments may induce a large flux of organic matter to the sea bottom. The organic carbon from the sediment in our study area showed a landward increase (see Table 2) suggesting that it did not originate from a marine environment [23].

3.2. Bottom sediment metals

3.2.1. Analytical quality assurance

The accuracy of the digestion method was appraised using standard reference material CRM 320 (river sediment of known elemental concentration) and subjecting this material to the same analytical procedure as the samples. Mean experimental and certified values, both with their standard deviations, are here presented in Table 3. A good agreement was obtained for most metals.

Table 2. Mean (n = 5) and percentage concentrations of the carbon content distribution in sediment sample from western and eastern Scheldt.

Station	TC content $(mg \cdot g^{-1})$	$\begin{array}{c} \text{OC content} \\ (\text{mg} \cdot \text{g}^{-1}) \end{array}$	% OC content	IC content $(mg \cdot g^{-1})$	% IC content
1	2.61 (1.47)	0.22 (0.12)	8.43	2.39 (1.47)	91.57
2	3.95 (1.71)	0.37 (0.08)	9.37	3.58 (1.66)	90.63
3	2.60 (0.74)	0.18 (0.08)	6.92	2.43 (0.73)	93.08
4	13.71 (3.84)	4.25 (1.69)	30.99	9.46 (2.21)	69.01
5	51.38 (7.36)	37.35 (5.93)	72.69	14.03 (6.71)	27.31
6	21.19 (4.20)	5.66 (3.60)	26.71	15.54 (4.28)	73.29
7	6.44 (0.34)	2.29 (0.72)	35.56	4.15 (0.73)	64.44

Notes: Numbers in parentheses represent standard deviations. TC, total carbon; OC, organic carbon; IC, inorganic carbon.

Metal	Reference standard materials Experimental value (SD)	Reference standard materials Certified value (SD)		
Fe	39,200 (39)	[45,000]*		
Cr	69.85 (8.77)	75.2 (10.42)		
Zn	154.92 (9.63)	142.00 (8.23)		
Cu	52.65 (2.45)	44.1 (1.86)		
Pb	51.83 (7.71)	42.3 (4.61)		
Cd	0.88 (0.18)	0.533 (0.06)		
Co	13.57 (0.48)	[19.00]*		

Table 3. Mean experimental values for the CRM 320 river sediment reference standard material calculated from six replicates with standard deviations and certified values.

Notes: (Concentrations are in $\mu g \cdot g^{-1}$ dry wt). Numbers in parentheses represent standard deviations *Noncertified, indicative values.

	Cd	Co	Cr	Cu	Pb	Fe	Zn
			F_1				
TC	0.99	0.89	ns	ns	-ve	-ve	0.83
OC	0.78	0.90	ns	ns	-ve	-ve	0.88
IC	ns	ns	ns	ns	-ve	-ve	ns
			F_2				
TC	0.96	0.94	0.98	0.94	0.94	0.95	0.96
OC	0.99	0.99	0.99	0.99	0.99	0.99	0.99
IC	ns	ns	ns	ns	ns	ns	ns
			F ₃				
TC	0.95	0.95	0.93	0.96	0.96	0.91	ns
OC	0.99	0.98	0.97	0.99	0.88	0.92	ns
IC	ns	ns	ns	ns	0.87	ns	ns
			Total met	al			
TC	0.93	0.95	0.94	0.95	0.96	0.92	Ns
OC	0.99	0.99	0.97	0.99	0.99	0.94	Ns
IC	ns	ns	ns	ns	ns	ns	ns
			Grain siz	e			
$<2 \mu m$	0.98	0.92	0.99	0.89	0.85	0.97	0.70
2–16 µm	0.74	0.80	0.92	0.64	0.76	0.86	0.60
16-63 µm	0.68	0.67	0.82	0.72	0.64	0.72	0.58
63–125µm	0.52	0.56	0.68	0.50	0.48	0.70	0.46
125–250 µm	ns	ns	ns	ns	ns	ns	ns
$250-500\mu m$	ns	ns	ns	ns	ns	ns	ns
500–100 µ m	ns	ns	ns	ns	ns	ns	ns

Table 4. Correlation between heavy metal fractions, gain size and organic carbon content of sediment.

Notes: TC, total carbon; OC, organic carbon; IC, inorganic carbon. ns, correlations not significant at p < 0.05.

3.2.2. Sequential extraction results for the bottom sediment

All metals extractable by acetic acid (F_1) correlated significantly with organic carbon, except Cr and Cu (p < 0.05) (Table 4). Although most of these correlations were positive, Pb and Fe showed significant negative correlation with organic carbon with all forms of carbon (total, organic and inorganic). All metals (except Zn) from F_1 showed a significant positive correlation with inorganic carbon. Metals from F_2 and F_3 correlated significantly with organic carbon indicating that organic matter is a good scavenger of heavy metals and can act as a vehicle for metal settling in a marine environment.

	Total concentration ($\mu g \cdot g^{-1}$)								
Fraction	Cd	Co	Cr	Cu	Pb	Fe	Zn		
1	0.04 (0.01)	0.59 (0.18)	115.53 (6.04)	5.41 (1.89)	3.77 (6.04)	3782.18 (6.04)	2.08 (0.72)		
2	0.01 (0.01)	0.93 (0.34)	186.9 (6.04)	3.35 (0.99)	5.39 (6.04)	4422.7(6.04)	8.23 (3.04)		
3	0.04 (0.01)	0.67 (0.28)	58.29 (6.04)	3.06 (1.21)	4.51 (6.04)	3850.1 (6.04)	6.91 (2.02)		
4	0.08 (0.01)	3.85 (0.98)	42.64 (6.04)	6.56 (1.84)	10.73 (6.04)	19290.78 (6.04)	45.92 (8.08)		
5	11.08 (3.15)	14.13 (4.56)	239.89 (6.04)	42.1 (16.01)	62.93 (6.04)	39380.86 (6.04)	144.97 (52.04)		
6	0.17 (0.04)	1.63 (1.50)	81.92 (6.04)	7.15 (2.86)	10.01 (6.04)	7149.01 (6.04)	13.94 (5.08)		
7	0.11 (0.04)	1.56 (0.84)	48.31 (6.04)	6.26 (2.64)	7.62 (6.04)	5891.86 (6.04)	23.27 (9.02)		

Table 5. Total metal concentration and their standard deviation (in parentheses) in bottom sediment obtained from Scheldt estuary.

3.2.3. Total metals in sediment

Sediments act as sinks for heavy metals and the degree of scavenging activity of sediments is a function of their physical, chemical and biological properties. It is common that heavy metals associate more with sediments of smaller grain size. In this study, a significant correlation (p < 0.05) was found for all metals between heavy metal concentrations in the bottom sediment and sediment grain sizes of 63–125 µm. Cadmium showed a significant correlation (p < 0.05) with sediment grain sizes > 125 µm. The levels of total metals in the sediment at all sites are presented in Table 5. Iron was the predominant metal at all sites, followed by Cr, Zn, Pb, Cu, Co and Cd.

3.2.4. F_1 -carbonate and exchangeable metals

Metals in this fraction are the most mobile and readily available for biological uptake in the environment. Metal associated with the carbonate and exchangeable fraction in contaminated sediments may reach up to 50% of the total metals [24]. In this study, metals were concentrated < 50%. The trend shows a high concentration of Cd followed by Co and Zn. The respective percentages of metals associated with the F₁ fraction varied between 2 and 38% (Cd), 2 and 11% (Co) and 1 and 17% (Zn) of total metal content. Other metals were present in this fraction in small concentrations (Table 6). The concentration of Cu, for example, was low, probably due to the poor adsorption behaviour of Cu on carbonate minerals. Cu has high affinity for dissolved organic matter and less affinity for adsorption on CaCO₃. Carbonates are also regarded as inefficient metal-carrier phases. The absence and/or presence of metals at low concentrations at most sampling stations may indicate the poor availability of these elements to benthic organisms.

3.2.5. F_2 -metals bound to organic matter

Heavy metals have high affinity for organic matter, the decomposition product of which can be appreciated for the behaviour of heavy metals in aquatic environments. Organic matter plays a key role in the scavenging, distribution and dispersion of heavy metals in the environment through cation exchange and chelation mechanisms. In this study, the heavy metal concentration in the organically bound fraction was appreciable for most metals at sampling sites 4–6; Pb being more important at these sites followed by Cd, Cu and Zn. Toxic metals such as Cd, Cu and Pb were partitioned in this fraction at between 6 and 72% (Cd), 1 and 47% (Cu) and 16 and 83% (Pb) (Table 6). These results show a behaviour similar to the results of metal burdens in the tissue of *M. edulis*, as studied by Shilla et al. [25]. F_1 and F_2 metal fractions are considered the most labile, can easily be leached and are therefore available for uptake by organisms.

Fraction	Site	Cd	Co	Cr	Cu	Pb	Fe	Zn
	1	14.39	11.67	1.64	1.22	1.66	0.72	1.13
	2	38.34	4.46	0.50	0.31	1.15	1.44	1.61
	3	20.83	9.11	8.88	1.03	1.68	0.98	4.26
F ₁	4	22.54	5.32	0.02	0.58	0.69	0.32	16.79
	5	4.14	2.40	0.01	0.15	0.02	0.01	6.67
	6	2.19	5.67	0.22	0.49	0.15	0.04	3.04
	7	23.40	6.46	0.17	0.65	1.75	1.85	17.12
	1	14.45	8.29	3.76	1.55	16.03	3.81	0.78
	2	24.62	4.52	1.49	1.16	21.04	5.86	0.38
	3	6.16	5.01	2.34	2.29	35.23	7.50	2.66
F_2	4	50.31	6.20	1.25	20.03	56.01	7.15	17.21
	5	72.78	24.04	3.08	47.13	83.48	16.23	42.50
	6	62.55	10.62	7.42	10.20	25.85	8.45	17.35
	7	32.59	8.33	1.07	12.39	33.15	9.30	9.35
	1	71.16	80.04	94.60	97.24	82.31	95.47	98.09
	2	37.03	91.02	98.01	98.53	77.81	92.71	98.01
	3	73.00	85.88	88.77	96.69	63.10	91.53	93.07
F ₃	4	27.15	88.48	98.73	79.39	43.30	92.53	65.99
	5	23.09	73.56	96.92	52.72	16.50	83.77	50.82
	6	35.27	83.71	92.36	89.31	74.01	91.51	79.61
	7	44.01	85.21	98.77	86.96	65.10	88.85	73.53

Table 6. Percentages of metal concentration in the three sedimentary fractions obtained by sequential extraction on bottom sediment samples.

3.2.6. F_3 -residual fraction (metals associated with the crystal lattice of silicates)

This fraction represents the part of metals that is largely unavailable to organisms. Residual metal fractions are not soluble under experimental conditions because they are held within the mineral matrix and their major input to the marine environment is not via anthropogenic sources. This fraction includes major transition metals whose concentrations are controlled by the mineralogy and weathering of the rocks in aquatic environments. In other words, this fraction is important because it gives a better insight into mechanism by which heavy metals are associated with mineralogical phases of the sediments. The results of this study indicate that at almost all of the sampling stations > 50% of the total metals are located in the residue fractions. Fe, Zn, Cu, Cr and Pb constituted 83–95, 50–98, 52–98, 88–98 and 16–82% of the total metal concentration respectively. The percentage of other metals varied between 73–91% (Co) and 23–71% (Cd).

4. Discussion

4.1. Grain size distribution and the carbon content of the sediment

As expected, the concentration of heavy metals showed a dependent relationship with the grain size of the sediment. Heavy metals were highly concentrated in sediments with smaller grain size, possibly due to the surface properties of clay minerals. Finer grains also have a larger surface area to volume ratio which enhances metal binding [26]. A high carbon content in the sediment also indicated a relationship with the total metals in the sediment, meaning that carbon, and in particular organic carbon, has a high affinity to metals, sequestering and storing them in a matrix. The correlation coefficients for the various metals to the total carbon content of the sediment at p = 0.05 are shown in Table 4. Both total metal concentrations in the sediment and the sequentially extracted fractions F_1 and F_2 correlated well with grain sizes up to $125 \,\mu$ m (Table 4). The sediment

grain size and its carbon content are vital parameters in the partitioning of metals in the sediment. Because the sediment particles are crucial as far as the selective behaviours of filter feeders are concern, metals bound to the active sites of sediments, together with those sequestered into the matrix of organics, can be taken up, assimilated and incorporated into the tissues of biota, which may be of public concern.

4.2. Total metals in sediment

Sediment is an important pool for heavy metals in marine systems. When the marine environment is in a dynamic state, there is an interaction between the sedimentary phase, the dissolved phase and the particulate phase through processes like resuspension, settling, adsorption and desorption. Hence sediment may play a key role as an indirect metal uptake route for organisms. Total metal concentrations of Co, Cu, Fe, Pb and Cr in sediment showed a landward increase at all sites. These metals showed a significant correlation (p < 0.05) with sediment grain size as well as sediment carbon content, which also showed a landward increase.

4.3. Metal speciation in sediments

Sequential extraction can be regarded as a procedure in which the natural sediment is transferred to a well-defined artificial environment and the shift in equilibria is registered. Therefore, sequential extraction can be used as an effective tool to reduce the complexity of the natural system and to gain a complete understanding of trace metal speciation and mobility in sediments. The exchangeable/mobile fraction is the most important because this portion is a candidate to resupply elements to the water body. Although the mobile fraction in this study correlated well with organic carbon and sediment grain size, its concentration was lowest compared with the F₂ and F_3 fractions. Both acetic acid-extracted (F_1) and EDTA-extracted (F_2) metals showed a similar trend at all sites. Only Cd, Co and Zn were extracted in $F_1 > 10\%$ in relation to their total content. Metals in F₂ were extracted up to 72% (Cd), 24% (Co), 47% (Cu), 83% (Pb), 16% (Fe) and 42% (Zn) for samples from site 5. Organic matter, as stated earlier, is a scavenger of metals and metals, in turn, preferentially associate with organic matter. Most metals were retained in the residual, the per cent extraction of which was 23-71 (Cd), 73-91 (Co), 88-98 (Cr), 52-98 (Cu), 16-82 (Pb), 83–95 (Fe) and 50–98 (Zn). The increase in metal concentrations in F_1 , F_2 , and F_3 was reflected by the increase in total metal concentrations in the sediment. Bhupendra [27] reported a similar trend in his study of metal mobility in sediments from Nepal.

The lower concentration of some metals extracted from the sediment into F_1 and F_2 may indicate lower exposure doses for marine organisms because the bioavailability of such metals to organisms is limited under normal conditions. Unless a special condition prevails in such marine sediments, the contribution from sediment alone may not be significant for the metals accumulated in the tissues of biota. Increased solubility of these metals, for example, may result in an increased leaching rate and exchangeable or biologically available pools of metals in the sediment. The solubility of Pb, for example, starts to increase at lower pH values (4 or lower), as reported by Bhupendra [27]. Although the amount of Pb extracted by EDTA was appreciable, the pH of the extracting solution (7) may still have had an effect. In general, the lower the pH of the sediment in the environment the higher the mobility (bioavailability) of Pb; the toxicity may be higher also. The availability of the nonessential element Cd and the essential element Zn (F_2 fraction) was better, probably because of the higher pH of the extracting solution, which was close to the pH at which Cd and Zn solubilises (6.5 for Cd and 6.0 for Zn).

A simple sequential extraction procedure, as presented in this study, has elucidated the speciation of heavy metals in the sedimentary phase and their relationship with organic matter content and sediment grain size. As mentioned earlier, the role of sediment in metal speciation in the marine environment is not insignificant. Animals such as filter feeders are secondarily or indirectly exposed to metals present in sediment and therefore for them, it is not a direct uptake route. In dynamic situations, the sediment compartment may interact with the organism's main metal uptake routes, namely, dissolved and particulate phases, and be exposed thereto.

Acknowledgements

We are very thankful to the Belgian Government for financing this work through the VLIR Scholarship. We are also thankful to Prof. Blust and team (Department of Biology, University of Antwerp, Belgium) for their help, advice and support while conducting this study.

References

- M.R. Lasheen and N.S. Ammar, Speciation of some heavy metals in River Nile sediments, Cairo, Egypt, Environmentalist 29 (2009), pp. 8–16.
- [2] C. Nicoletta, M.P. Bianca, and P. Marco, Metal bioavailability: how does its significance change in the time? Ann. Chim. 96(3–4) (2006), pp. 131–136.
- [3] Z.S. Ahnstrom and D.R. Parker, Development and assessment of a sequential extraction procedure for the fractionation of soil cadmium, Soil Sci. Soc. Am. J. 63 (1999), pp. 1650–1658.
- [4] N.J. Hu, Z.Q. Li, P. Huang and T. Cheng, Distribution and mobility of metals in agricultural soils near a copper smelter in South China, Environ. Geochem. Health 28 (2006), pp. 19–26.
- [5] S. Ray, D.W. Mcleese, and M.R. Peterson, Accumulation of copper, zinc, cadmium and lead from two contaminated sediments by three marine invertebrates. A laboratory study, Bull. Environ. Contam. Toxicol. 26 (1981), pp. 315–322.
- [6] J.H. Rule and R.W. Alden III, Cadmium bioavailability to three estuarine animals in relationship to geochemical fractions in sediments, Arch. Environ. Contam. Toxicol. 19 (1990), pp. 878–885.
- [7] O. Abollino, M. Aceto, C. Sarzanini, and E. Mentasti, The retention of metal species by different solid sorbents: mechanisms for heavy metal speciation by sequential three-column uptake, Anal Chim Acta 411 (2000), pp. 223–237.
- [8] K. Polyák and J. Hlavay, Environmental mobility of trace metals in sediments collected in the lake Balton, Fresenius J. Anal. Chem. 363 (1998), pp. 587–593.
- [9] J.L. Gómez-Ariza, I. Giráldez, D. Sanchez-Rodas and E. Morales (2000). Comparison of the feasibility of three extraction procedures for trace metal partitioning in sediments from south-west Spain, Sci. Total Environ. 246 (2000), pp. 271–283.
- [10] H. Akcay, A. Oguz, and C. Karapirec, Study of heavy metal pollution and speciation in Buyak Menderes and Gediz river sediments, Water Res 37 (2003), pp. 813–822.
- [11] S.P. Singh, F.M. Tack, and M.G. Verloo, *Heavy metal fractionation and extractability in dredged sediment derived surface soils*, Water, Air Soil Pollut. 1021 (1998), pp. 313–328.
- [12] Z. Wang, X.-q. Shan, and S. Zhang, Comparison of speciation and bioavailability of rare earth elements between wet rhizosphere soil and air-dried bulk soil, Anal. Chim. Acta 441 (2001), pp. 147–156.
- [13] D. Panda, V. Subramanian, and R.C. Panigrahy, Geochemical fractionation of heavy metals in Chilka Lake (east coast of India) – a tropical coastal lagoon, Environ. Geol. 26 (1995), pp. 199–210.
- [14] S.M.N. Moalla, R.M. Awadallah, M.N. Rashed, and M.E. Soltan, Distribution and chemical fractionation of some heavy metals in bottom sediments of Lake Nasser, Hydrobiologia, 364(1) (1997), pp. 31–40.
- [15] R. Rosental, G.A. Eagle, and M.J. Orren, The trace metal distribution in different chemical fractions of nearshore marine sediments, Estuar. Coast. Shelf Sci. 22 (1986), pp. 303–324.
- [16] C. Carapeto and D. Purchase, Use of sequential extraction procedure for the analysis of cadmium and lead in sediment samples from a constructed wetland, Bull. Environ. Contam. Toxicol. 64 (2000), pp. 51–58.
- [17] A. Tessier, P.G.C. Campbell, and M. Bisson, Sequential extraction procedure for the speciation of particulate trace metals, Anal. Chem. 51 (1979), pp. 844–851.
- [18] C.N. Mulligan, R.N. Yong, and B.F. Gibbs, An evaluation of technologies for the removal of heavy metals from sediments, J. Hazard. Mater. 85 (2001), pp. 145–163.
- [19] L.S. Ngiam and P.E. Lim, Speciation patterns of heavy metals in tropical estuarine anoxic and oxidized sediment by different sequantional extraction schemes. Sci. Total Environ. 275 (2001), pp. 53–61.
- [20] W. Baeyens, G.T.M. Van Eck, C. Lambert, R. Wollast, and L. Goeyens, *General description of the Scheldt estuary*, Hydrobiologia. 366 (1998), pp. 1–14.
- [21] D.-J. Whitworth, E.P. Achterberg, V. Herzl, M. Nimmo, M. Gledhill and P.J. Worsfold, Development of a simple extraction procedure using ligand competition for biogeochemically available metals of estuarine suspended particulate matter, Anal. Chim. Acta. 392 (1999), pp. 3–17.
- [22] R. Van Ryssen, M. Leermakers, and B. Baeyens, *The mobilization potential of trace metals in aquatic sediments as a tool for sediment quality classification*, Environ. Sci. Policy 2 (1999), pp. 75–86.
- [23] E. Ruiz, F. Romero, and G. Besga, Selective solubilization of heavy metals in torrential river sediments, Toxicol. Environ. Chem. 33(1) (1991), pp. 1–6.

- [24] Y. Liang and M.H. Wong, Spatial and temporal organic and heavy metal pollution at Mai Po Marshes Nature Reserve, Hong Kong, Chemosphere 52 (2003), pp. 1647–1658.
- [25] D. Shilla, D. Qadah, and M. Kalibbala, Distribution of heavy metals in dissolved, particulate and biota in the Scheldt estuary, Belgium, Chem. Ecol. 24(1) (2008), pp. 1–14.
- [26] V.C. Lakhan, K. Cabana, and P.D. LaValle, *Relationship between grain size and heavy metals in sediments from beaches along the coast of Guyana*, J. Coastal Res. 19(3) (2003), pp. 600–608.
- [27] D. Bhupendra, Total and extractable (mobilizable and mobile) Heavy metals in the Bagmat River sediment of Kathmandu, Nepal, J. Environ. (Nepal). 6(7) (2001), pp. 34–49.